

Evaluation of ESwab (COPAN) for the detection of *Ureaplasma urealyticum* and *Mycoplasma hominis* from genital specimens with Mycoplasma Duo kit (BIO-RAD)

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OBJECTIVES :

To validate ESwab as an alternative collection and transport medium for identification and titration of genital mycoplasma by using the Mycoplasma Duo kit

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METHODS :

1) Comparison between 3 different methods for Mycoplasma culture, identification and titration of 25 clinical specimens and various dilutions of *U. parvum* ATCC 27815 (*Up*) and *M. hominis* ATCC 27618 (*Mh*) strains :

- Method 1** : collection by a Rayon swab, transfer into the suspension medium of the Mycoplasma Duo kit (Bio-Rad France) and microplate seeding according to manufacturer's instructions
- Method 2** : collection and transport by Eswab (Copan, Italy) and transfer of 200 µl of ESwab medium into the suspension medium of the Mycoplasma Duo kit before microplate seeding
- Method 3** : collection by ESwab and direct seeding of the microplate by 100µl of Eswab medium

- Transport < 48 hours at room temperature
- Inoculation of an A7 agar (BioMérieux, France) as gold standard, in each case
- Incubation, reading and interpretation according to manufacturer's instructions

2) Statistical analysis of patient's results during one year before and after introducing Eswab for mycoplasma diagnosis in our lab (comparing percentages)

RESULTS :

ATCC strains

Results	Method 1		Method 2		Method 3	
	Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar
<i>U. parvum</i> ATCC 27815						
Dilution 1/10	≥10 ⁴	10 ⁴	≥10 ⁴	10 ⁴	≥10 ⁴	10 ⁵
Dilution 1/20	≥10 ⁴	10 ³	≥10 ⁴	10 ³	≥10 ⁴	10 ⁵
Dilution 1/100	≥10 ⁴	10 ⁴	≥10 ⁴	10 ³	≥10 ⁴	10 ⁴
Dilution 1/1000	≤10 ³	10 ³	≤10 ³	10 ³	∅	∅

Results	Method 1		Method 2		Method 3	
	Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar
<i>M. hominis</i> ATCC 27618						
No dilution	≥10 ⁴	10 ⁵	≥10 ⁴	10 ⁵	∅	10 ³
Dilution 1/10	≤10 ³	10 ³	≥10 ⁴	10 ³	∅	∅
Dilution 1/100	≤10 ³	10 ³	≤10 ³	10 ³	∅	∅

With ATCC strains, there was no difference between the method 1 and 2. The method 3 didn't allow the detection of *Mh* at every concentration and *Up* at the lowest.

Clinical specimens

About the 25 clinical specimens :

- 17 negative cultures with every method
- 2 samples contaminated by yeast
- 3 positive cultures with *U. urealyticum* (*Uu*) ≥ 10⁴ colour changing unit/ml for the 3 methods
- 1 culture positive with *Uu* ≤ 10³ CCU for methods 1 and 2 and negative for method 3
- 1 culture positive with *Uu* ≥ 10⁴ CCU for the 3 methods and *Mh* ≥ 10⁴ CCU for methods 1 and 3 and *Mh* ≤ 10³ CCU with method 2
- 1 culture positive with *Uu* ≥ 10⁴ CCU for methods 1 and 3 and *Uu* ≤ 10³ CCU with method 2

Results	Specimen	Method 1		Method 2		Method 3	
		Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar	Mycoplasma Duo kit	A7 Agar
Patient 1	Urethral	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> < 10 ³	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵
		<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅
Patient 2	Vaginal	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵
		<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅
Patient 3	Vaginal	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁶	<i>Uu</i> ≤ 10 ³	<i>Uu</i> 10 ⁵	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁶
		<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅
Patient 4	Vaginal	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁴	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> 10 ⁵
		<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅
Patient 5	Vaginal	<i>Uu</i> ≤ 10 ³	<i>Uu</i> ≤ 10 ³	<i>Uu</i> ≤ 10 ³	<i>Uu</i> ≤ 10 ³	<i>Uu</i> ∅	<i>Uu</i> ≤ 10 ³
		<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅	<i>Mh</i> ∅
Patient 6	Vaginal	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> ≥ 10 ⁴	<i>Uu</i> ≥ 10 ⁴
		<i>Mh</i> ≥ 10 ⁴	<i>Mh</i> ≤ 10 ³	<i>Mh</i> ≤ 10 ³	<i>Mh</i> ≤ 10 ³	<i>Mh</i> ≥ 10 ⁴	<i>Mh</i> ≤ 10 ³

Results of the six positive clinical specimens

Statistical analysis of patient's results during one year before and after introducing Eswab as collection medium

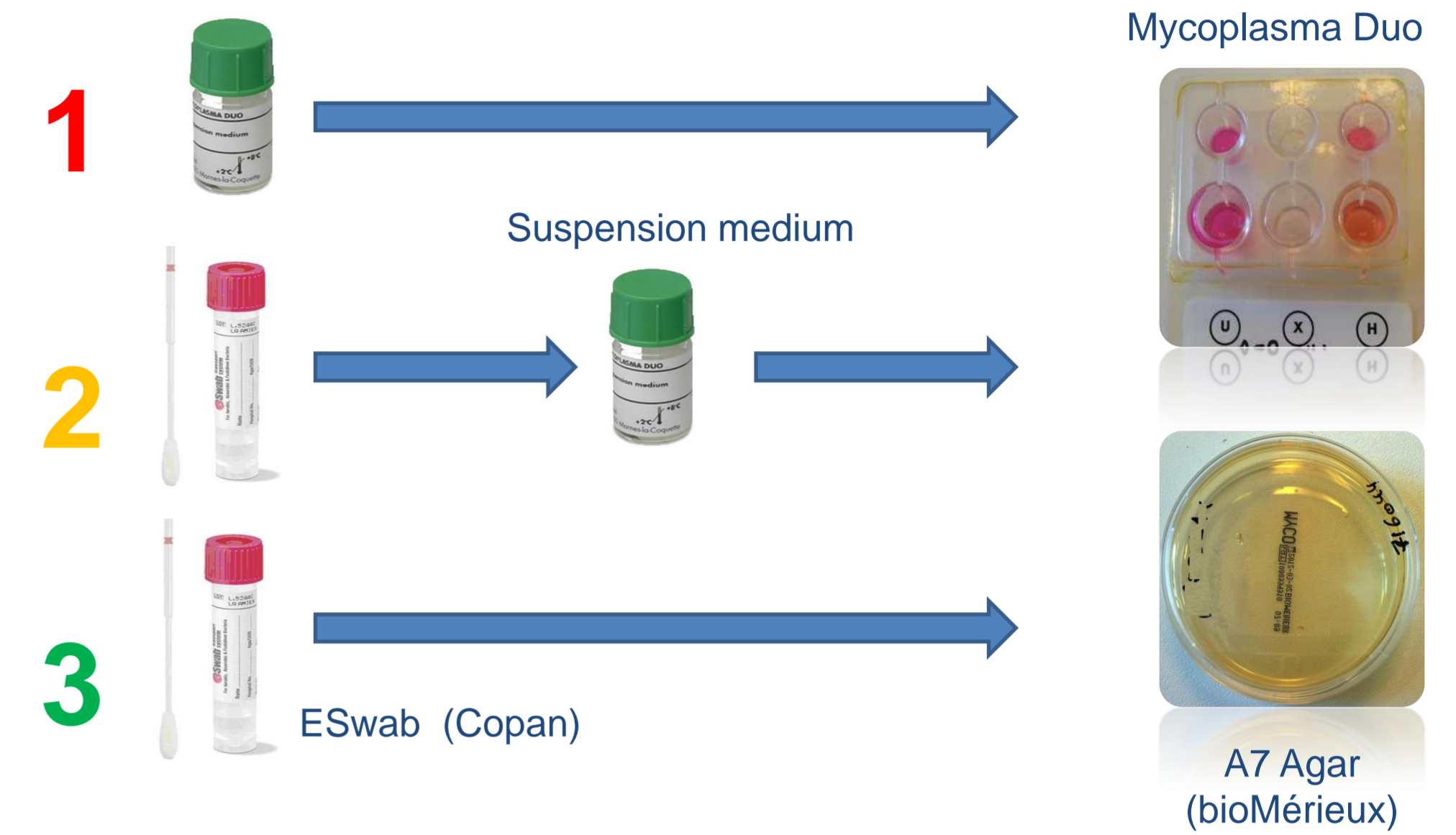
No statistically significant difference between the two study periods was found : 269 specimens (27%) positive in 2009 versus 320 (26%) in 2010 with about a thousand clinical specimens in each cohort

Year	2009	2010	Year	2009	2010
<i>Uu</i> negative	729 (73%)	918 (74%)	<i>Mh</i> negative	960 (96%)	1200 (98%)
<i>Uu</i> ≤ 10 ³	61 (6%)	62 (5%)	<i>Mh</i> ≤ 10 ³	16 (1,6%)	62 (0,9%)
<i>Uu</i> ≥ 10 ⁴	208 (21%)	258 (21%)	<i>Mh</i> ≥ 10 ⁴	22 (2,2%)	27 (2,2%)
TOTAL	998	1238	TOTAL	998	1238

Comparison of *U. urealyticum* and *M. hominis* results between 2009 and 2010

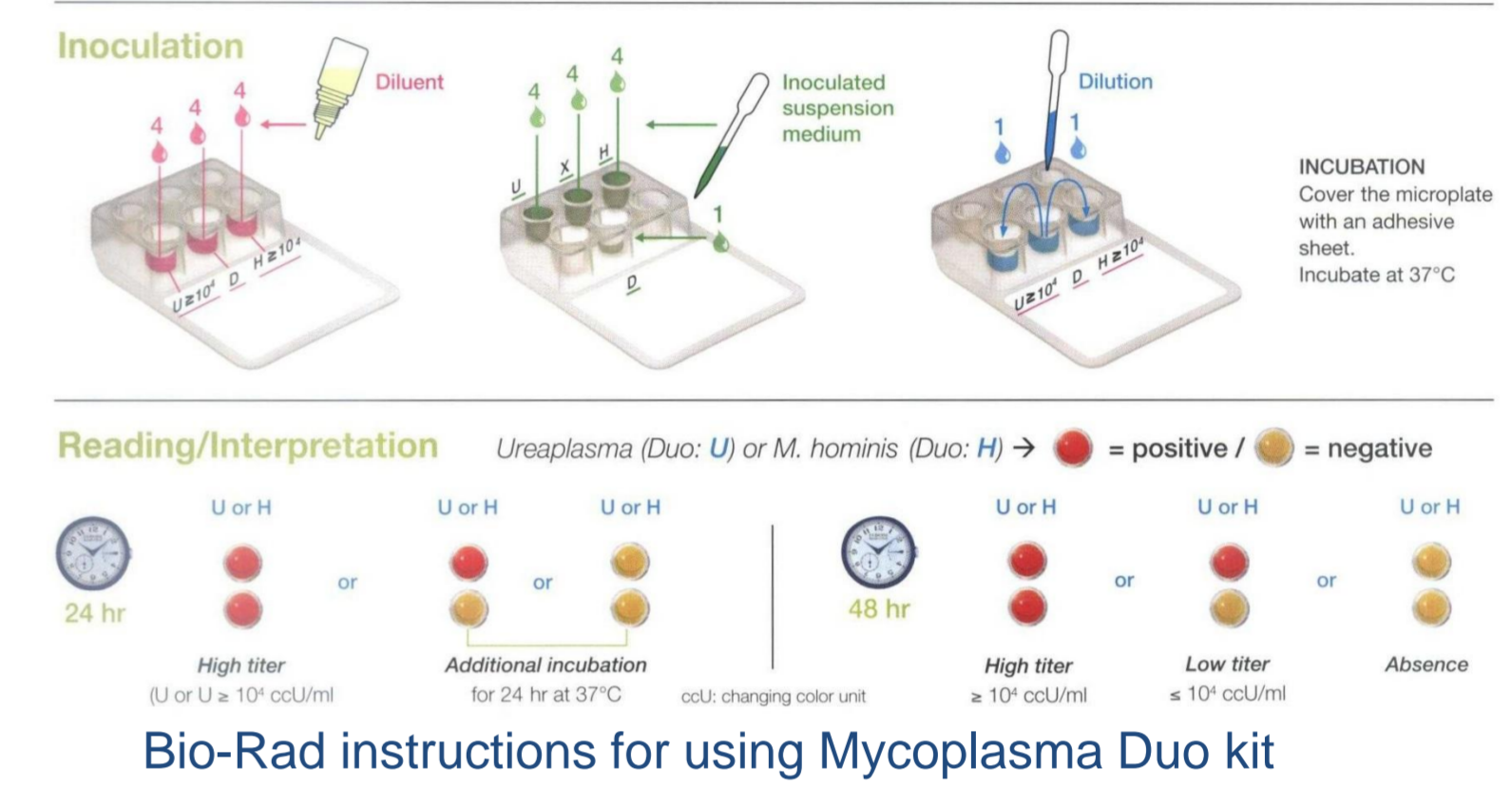
CONCLUSION :

- Transfer of 200 µl ESwab into the Bio-Rad suspension medium** provides equivalent results in comparison with the method recommended by Bio-Rad.
- Use of ESwab** for collection and transport of mycoplasma with Mycoplasma Duo kit **simplifies the detection of genital mycoplasma** by allowing multiple testing from the same original specimen.



Mycoplasma DUO: identification, titration of *Ureaplasma* and *M. hominis*

Specimen: Swabs collected by scraping mucosa (urethra, cervix...) and then soaked in suspension/transport medium, can be kept 48 hours at room temperature or 72 hours at +2-8°C.



Bio-Rad instructions for using Mycoplasma Duo kit