# Stability of *Bordetella pertussis* and *Bordetella parapertussis* in the ESwab Transport System for Culture and PCR

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## **Revised Abstract**

#### **Objectives**

Pertussis (whooping cough) is caused by *Bordetella pertussis* and *Bordetella parapertussis*, with several states in the USA reporting an increased incidence. Laboratory detection methods include culture, direct fluorescence assay, and PCR methods; successful culture and PCR detection require proper specimen collection and transport. This study examined the effectiveness of the ESwab system (consisting of liquid Amies transport medium and a flocked nasopharyngeal swab, Copan Diagnostics, Inc.) for maintenance of viability of *B. pertussis* and *B. parapertussis* for culture and preservation of nuclear material for detection by PCR.

#### Methods

Eight Bordetella isolates were tested, including 5 B. pertussis isolates (ATCC 9340 and 8467 strains, and 3 recent clinical isolates) and 3 B. parapertussis isolates (ATCC 15237 strain and 2 recent clinical isolates). Test methods were based on CLSI M40 guidelines. Three Bordetella saline suspensions (108, 106, and 10<sup>4</sup> CFU/mL) were prepared. Each suspension was used to inoculate ESwabs in triplicate. The inoculated ESwabs were stored refrigerated (2-8°C) for up to 96 hours prior to plating. Bacteria from the ESwabs were cultured on Regan Lowe agar plates at 0, 24, 48, and 96 hours of refrigerated storage postinoculation; plates were then incubated at 3-7°C in ambient air for a minimum of 4 days. The numbers of colonies at 24, 48, and 96 hours were compared to the 0 hour count to determine percent recovery (viability). Real-time PCR, using primers targeting the IS481 gene of B. pertussis and the IS1001 gene of B. parapertussis, was performed on 24-hour and 96-hour inoculum in the ESwab.

### Results

Bordetella was isolated from all of the ESwabs after 96 hours of refrigerated storage. Percent recovery ranged from 27.2% to 96.3% for *B. pertussis*, and from 32.7% to 74.5% for *B. parapertussis*, and was similar across the three inoculum densities for all isolates tested. PCR detected *B. pertussis* and

B. parapertussis in all of the 24- and 96-hour ESwabs, regardless of initial inoculum concentration ( $10^4$  or  $10^6$ ).

#### Conclusion

The ESwab maintains sufficient viability of *B. pertussis* and *B. parapertussis* to permit detection in bacteria cultures and preserves DNA integrity for PCR detection, even after 96 hours of refrigerated storage.

## Introduction

Infection with *Bordetella pertussis*, the bacterium that causes pertussis (whooping cough), is spread by person-to-person transmission via aerosolized respiratory droplets or by direct contact with respiratory secretions. Pertussis manifests with mild upper respiratory tract symptoms that begin 7–10 days (range 6–21 days) after exposure, followed by a severe lingering cough

## Introduction (cont)

that becomes paroxysmal and can last for weeks or even months. Coughing paroxysms vary in frequency and are often followed by vomiting. A similar, milder disease is caused by *B. parapertussis*.

The United States is currently experiencing a resurgence of pertussis, most notably in California where a total of 1,337 cases were reported during the period of January 1 to June 30, 2010. This represents a 418% increase from the same period in 2009. Part of this increase may reflect the natural cycle of pertussis, which exhibits incidence peaks approximately every 5 years; 2010 was at the peak of the cycle. Other contributing factors may include a lower degree of immunity from the current DTaP vaccine relative to the older DTP vaccine, waning immunity in older children and adults, and low rates of booster shots to maintain immunity.

According to current CDC guidelines, laboratory methods for confirmation of pertussis cases meeting the clinical case definition include culture and PCR. Specimens for culture and PCR are often collected from the nasophayngeal region by swab. Other swab systems may not maintain viability of *Bordetella* without added charcoal, and charcoal swabs may not be acceptable for PCR. This study examined the effectiveness of the ESwab system (consisting of liquid Amies transport medium and a flocked nasopharyngeal swab, Copan Diagnostics, Inc.) for maintenance of viability of *B. pertussis* and *B. parapertussis* for culture and preservation of nuclear material for detection by PCR.

## Methods

A total of 8 strains of *Bordetella* were tested. Five strains (3 *B. pertussis* and 2 *B. parapertussis*) were recent clinical isolates obtained from Quest Diagnostics Nichols Institute (San Juan Capistrano, CA) or Focus Diagnostics. Three quality-control strains were tested including *B. pertussis* ATCC 8467 (Hardy Diagnostics), *B. pertussis* ATCC 9340 (Focus stock strain), and *B. parapertussis* ATCC 15237 (Focus stock strain).

## **Bacteria viability**

All tests for bacterial viability were modified from the quantitative elution method described in CLSI M40 A. Modifications included:

- 1. Inoculum suspensions of  $10^8$ ,  $10^6$ , and  $10^4$  CFU/mL were prepared and used for testing.
- 2. ESwabs were rolled into the inoculum, allowed to absorb for 10-15 sec, and then placed into 1 mL of liquid Amies transport medium.
- 3. The 108 and 106 CFU/mL ESwabs were processed as follows:
- a. The ESwab transport tube was considered the initial dilution tube (total of 10<sup>7</sup> CFU/mL) and was vortexed in the transport tube for 10-15 sec.
- b. 10-fold dilutions were prepared in 0.9 mL of sterile saline to reach a final concentration of 10<sup>3</sup> CFU/mL.

# Methods (cont)

4. The 10<sup>4</sup> CFU/mL ESwab remained undiluted.

The average CFU for each incubation time was recorded and compared to the average CFU at 0 h. A percent viability (average CFUs at a particular incubation time compared to the CFUs at 0 h) was obtained for each isolate.

## PCR Testing

Performed on all isolates at 0 hr and 96 hr with the 10<sup>6</sup> and 10<sup>4</sup> CFU/mL ESwabs. The 10<sup>8</sup> swabs were not tested.

- 1. Detection of *B. pertussis* and *B. parapertussis* by real-time PCR was accomplished using an in-house developed assay.
- 2. Nucleic acids were isolated from patient specimens using the Roche MagNA Pure LC system.
- 3. Real-time PCR was accomplished using primers and probes designed to hybridize to sequences found in *IS481* and *IS1001*.

## Results

- 1. Bordetella was isolated from all of the ESwabs after 96 hours of refrigerated storage (Table 1).
  - a. Percent recovery for *B. pertussis* ranged from 27.2% to 96.3%.
  - b. Percent recovery for *B. parapertussis* ranged from 32.7% to 74.5%.
  - c. Percent recovery was similar across the 3 inoculum densities for all isolates tested.
  - d. See Figure 1 for typical 0 hr vs 96 hour culture recovery.
- 2. PCR detected *B. pertussis* and *B. parapertussis* in all of the 24- and 96-hour ESwabs, regardless of initial inoculum concentration (10<sup>4</sup> or 10<sup>6</sup>). The 10<sup>8</sup> CFU/mL ESwabs were not tested. Table 2.

Table 1. Recovery of *Bordetella* strains from ESwab after 96 hr refrigeration

remger	renigeration												
Inoc.	0 hr	96 hr	% Recovery	Inoc.	0 hr	96 hr	% Recovery						
B. para	ssis Al	TCC 15237	B. pertussis strain Q2										
10 <sup>8</sup>	71	53	73.9	10 <sup>8</sup>	329	250	76.0						
10 <sup>6</sup>	57	38	67.3	10 <sup>6</sup>	208	160	76.9						
10 <sup>4</sup>	53	40	74.5	10 <sup>4</sup>	196	76	38.8						
B. para	ssis sti	rain Q3	B. pertussis strain Q1										
10 <sup>8</sup>	223	123	55.1	10 <sup>8</sup>	328	316	96.3						
10 <sup>6</sup>	222	141	63.5	10 <sup>6</sup>	318	214	68.3						
10 <sup>4</sup>	251	82	32.7	10 <sup>4</sup>	248	189	76.2						
	s <i>si</i> s stı	rain #20	B. pertussis Focus Isolate										
10 <sup>8</sup>	107	68	64.5	10 <sup>8</sup>	96	35	36.6						
10 <sup>6</sup>	152	104	68.4	10 <sup>6</sup>	41	29	69.5						
10 <sup>4</sup>	136	80	58.8	10 <sup>4</sup>	46	30	63.7						
B. peri	9340	B. pertussis ATCC 8467											
10 <sup>8</sup>	56	37	66.7	10 <sup>8</sup>	71	32	42.4						
10 <sup>6</sup>	50	39	77.5	10 <sup>6</sup>	71	28	39.2						
10 <sup>4</sup>	47	33	70.0	10 <sup>4</sup>	90	25	27.2						

Values are CFU/mL recovered at the final dilution.

# Results (cont)

Table 2. Bordetella PCR detection (cycles) from ESwab after 96 hr refrigeration

	Eswab							
Inoc.	#1	#2	#3	Inoc.	#1	#2	#3	
B. paraperti	ussis AT	CC 1523	B. pertussis strain Q2					
10 <sup>6</sup>	26.2	25.9	25.5	10 <sup>6</sup>	23.7	23.7	23.6	
10 <sup>4</sup>	32.1	33.9	33.0	10 <sup>4</sup>	30.7	31.1	30.9	
B. paraperti	ussis str	ain Q3	B. pertussis strain Q1					
10 <sup>6</sup>	25.0	25.2	NT	10 <sup>6</sup>	22.7	22.7	23.0	
10 <sup>4</sup>	26.0	26.00	25.6	10 <sup>4</sup>	26.0	25.9	26.3	
B. paraperti	ussis str	ain #20	B. pertussis -Focus Isolate					
10 <sup>6</sup>	24.2	23.8	23.8	10 <sup>6</sup>	21.4	20.5	21.2	
10 <sup>4</sup>	29.8	29.0	30.0	10 <sup>4</sup>	27.9	27.8	27.8	
B. pertussis	ATCC 9	340	B. pertussis ATCC 8467					
10 <sup>6</sup>	20.8	20.6	20.7	10 <sup>6</sup>	23.1	22.9	22.6	
10 <sup>4</sup>	26.0	26.1	26.2	10 <sup>4</sup>	26.2	25.9	25.5	
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Figure 1. *B. pertussis* strain Q2 recovery after sample refrigeration at (2°C to 8°C) for 0 hr (left) and 96 hr (right).

## Conclusions

- The ESwab in Amies liquid maintains sufficient viability of B. pertussis and B. parapertussis to permit detection in cultures after 96 hours of refrigerated storage.
- The ESwab in Amies liquid preserves DNA integrity for PCR detection after 96 hours of refrigerated storage.

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