Detection of *Streptococcus pneumoniae* from Different Types of Nasopharyngeal Swabs in Children

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Abstract

**Background:** A better understanding of the epidemiology of nasopharyngeal carriage of *Streptococcus pneumoniae* is important to assess the impact of vaccination and the pathogenesis of pneumococcal disease. We compared the recovery of *S. pneumoniae* from nylon flocked, Dacron and rayon swabs.

**Methods:** The recovery of *S. pneumoniae* from mock specimens using flocked, Dacron and rayon swabs were compared by culture. The yield from paired nasopharyngeal (NP) samples obtained from healthy children sampled with flocked and Dacron swabs was also determined using culture and lytA-targeted real-time polymerase chain reaction (qPCR).

**Results:** Using mock specimen, the percentage recovery of *S. pneumoniae* ATCC 49619 (serotype 19F) strain from the flocked swabs was 100%, while it was 41% from Dacron swabs and 7% from rayon swabs. Similar results were observed for *S. pneumoniae* serotypes 1 and 5. *S. pneumoniae* was cultured from 18 of 42 (43%) paired NP samples from the healthy children (median age 8 [interquartile range (IQR) 5–16] months). The median number of colony-forming units (CFU) recovered from flocked swabs was two-fold higher (8.8 × 10⁴ CFU/mL [IQR, 2.0 × 10³ – 4.0 × 10⁵ CFU/mL]) than Dacron swabs (3.7 × 10⁴ CFU/mL [IQR, 4.0 × 10³–3.2 × 10⁵ CFU/mL], p = 0.17). Using lytA-targeted qPCR from paired NP samples, the median copy number of *S. pneumoniae* detected from flocked swabs was significantly higher than from Dacron swabs (3.0 × 10⁵ genome copies/mL [IQR, 1.3 × 10⁴–1.8 × 10⁶] vs. 9.3 × 10¹⁶ genome copies/mL [IQR, 7.0 × 10⁶–1.1 × 10²²]; p = 0.005).

**Conclusion:** Flocked swabs released more *S. pneumoniae* compared to both Dacron and rayon swabs from mock specimens. Similarly, higher bacterial loads were detected by qPCR from flocked swabs compared with Dacron swabs from healthy children.

Introduction

*Streptococcus pneumoniae* is the leading bacterial cause of childhood pneumonia worldwide [1,2]. *S. pneumoniae* colonizes the nasopharynx of many healthy young children and only causes pneumonia in a small proportion of those colonized [3–6]. The timing of acquisition, intensity of colonization and interaction of *S. pneumoniae* with other respiratory pathogens are likely to be key determinants for progression to pneumonia [7–10]. Since nasopharyngeal (NP) colonization by *S. pneumoniae* is key to understanding the pathogenesis of pneumococcal disease and is increasingly used as an endpoint for pneumococcal vaccine studies, it is important to establish the optimal strategy for recovery of *S. pneumoniae* from NP specimens. Sampling of the NP may be achieved using an NP swab or aspirate. NP swabs are preferred as the procedure is simpler, quicker and better tolerated by children. The ideal swab would be highly absorbent, maintain the viability of microorganisms present, release most of the specimen material into culture broth or transport medium and not inhibit culture or nucleic acid amplification.

Traditionally used swabs, such as Dacron and rayon swabs are constructed by winding the respective fibres onto the tip of the swab shaft. This swab design may potentially trap a large proportion of clinical material in the fibre matrix, potentially inhibiting culture or nucleic acid amplification.

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There are limited data comparing the recovery of *S. pneumoniae* using different types of swabs [22]. Only one study has compared the recovery of *S. pneumoniae* by culture and nucleic acid amplification from Dacron, calcium alginate and rayon swabs [23]. No published study has compared the more recently available flocked swabs (Copan Italia, Brescia, Italy) with other swab types for the recovery of *S. pneumoniae*. Therefore, the aim of the present study was to compare the yield from different types of swabs using culture and nucleic acid-based detection methods.

**Materials and Methods**

**Pneumococcal recovery from mockspecimens using flocked, Dacron, and rayon NP swabs**

American Type Culture Collection (ATCC 49619) strain of *S. pneumoniae* (serotype 19F) as well as clinical isolates of *S. pneumoniae* serotypes 1 and 5 (donation from the Respiratory and Meningeal Pathogens Research Unit, National Institute for Communicable Diseases, National Health Laboratory Service, Johannesburg, South Africa) were sub-cultured onto Columbia blood agar base with 2% agar, 5% horse blood and 4 μg/mL gentamicin (CAG) (Greenpoint Media Laboratory, National Health Laboratory Service, Cape Town, South Africa) and incubated at 37°C in 5% CO₂ overnight. The resulting colonies were then inoculated into 10 mL Todd-Hewitt broth and inoculated at 37°C in 5% CO₂ for 3 hours to an optical density of 0.5 at 492 nm (which corresponds to an exponential phase of approximately 1.2×10⁹ CFU/mL, as determined previously [data not shown]).

Three different types of NP swabs were compared, the flocked swab (cat. no. 516C; Copan Italia, Brescia, Italy), Dacron swab (cat. no. MW151D; Medical Wire & Equipment, Corsham, United Kingdom) and rayon swab (cat. no. 160C; Copan Italia, Brescia, Italy). Log-phase cultures of each of the *S. pneumoniae* strains were serially diluted (10-fold) in phosphate buffered saline (PBS). For each of the *S. pneumoniae* strains, triplicate aliquots (20 μL) of the 10⁻²⁻¹⁰⁻³ dilutions were dispensed into 200 μL tubes. A single swab of each type was briefly placed into each of the triplicate aliquots of the above mentioned dilutions series until the inoculum was absorbed into the swab. This was performed for all swab types tested. The swabs were allowed to stand at room temperature for 10 minutes before transferring them into 1 mL of skim milk-tryptone-glucose-glycerol (STGG) transport medium.

The composition of the STGG medium used in this study is based on the triplicate aliquots of the above mentioned dilutions series until the inoculum was absorbed into the swab. This was performed for all swab types tested. The swabs were allowed to stand at room temperature for 10 minutes before transferring them into 1 mL of skim milk-tryptone-glucose-glycerol (STGG) transport medium. The resulting colonies were then inoculated into 10 mL Todd-Hewitt broth and inoculated at 37°C in 5% CO₂ for 3 hours to an optical density of 0.5 at 492 nm (which corresponds to an exponential phase of approximately 1.2×10⁹ CFU/mL, as determined previously [data not shown]).

Following sampling, swabs were immediately placed into 1 mL of STGG, transported on ice to the laboratory and frozen at −80°C for later batch processing. After thawing, STGG samples were vortexed for 15 seconds to disperse organisms from the swab. A 10 μL aliquot was then inoculated onto CAG media and incubated at 37°C in 5% CO₂ overnight.

**Total nucleic acid extraction from flocked and Dacron NP swabs from healthy children**

The STGG aliquots were thawed to room temperature (22°C) and vortexed for 15 seconds. Thereafter, 300 μL of each sample was subjected to automated total nucleic acid extraction on the QIAasymply SP instrument (Qiagen, Hilden, Germany) using the QIAsymphony® Virus/Bacteria mini kit (cat. no. 931036) according to the manufacturer’s instructions. Total nucleic acid was eluted in 60 μL elution buffer and stored at −20°C until processing.

**Identification of *S. pneumoniae* using lytA-targeted real-time PCR from NP samples collected in healthy children**

A quantitative real-time PCR (qPCR) assay for the detection of the *S. pneumoniae* autolysin-encoding gene (*lytA*) was performed to determine bacterial load recovered from either flocked or Dacron swabs collected from healthy children. Primers and probe used were those previously published by Carvalho et al. [29], and have been shown to be specific for *S. pneumoniae* detection [30]. The reaction mix contained 2.5 μL genomic DNA in a total reaction volume of 12.5 μL containing: 1×TaqMan® gene expression mastermix (Applied Biosystems, California, United States of America), 200 nM for each primer and probe. PCR amplification was performed on the Bio-Rad CFX96 Touch™ Real-Time PCR amplification system (Bio-Rad Laboratories, Hercules, CA, United States of America). The thermal cycling conditions consisted of an initial hot start of 30°C for 2 minutes, denaturation at 95°C for 10 minutes, followed by 40 amplification cycles of 95°C for 15 seconds, 60°C for 1 minute. The lowest limit of detection of the qPCR assay was 10 copies/μL as determined by inspection of a standard curve (10-fold serial dilution of *S. pneumoniae* ATCC 49619 strain genomic DNA). The quantification cycle (Cq) value at the detection limit point was 36. A positive (*S. pneumoniae* ATCC 49619) and non-template control (sterile water) were included in each run. Results below the lowest limit of detection were considered negative.

**Statistical analysis**

Statistical analysis was performed using GraphPad Prism version 6.01 (GraphPad Software Inc, California, United States of America) and STATA software version 11.0 (Stata Corporation, Texas, United States of America). For normally distributed data, our own findings (below) had demonstrated that flocked and Dacron swabs were better than rayon swabs for *S. pneumoniae* release [12,20,25–27], and there are practical difficulties in performing a 3-way comparison. Consecutive healthy children undergoing elective surgery at Red Cross War Memorial Children’s Hospital were enrolled. Swabs were obtained by a trained research nurse using a standardised procedure [28]. Paired NP swabs were obtained from each child from separate nostrils. A flocked swab was used to obtain a specimen from one nostril and a Dacron swab from the other nostril. The order of sampling was randomized. Written informed consent was obtained from a parent or legal guardian. The study was approved by the Human Research Ethics Committee of the Faculty of Health Sciences, University of Cape Town (HREC ref: 062/2011).

The STGG aliquots were thawed to room temperature (22°C) and vortexed for 15 seconds. Thereafter, 300 μL of each sample was subjected to automated total nucleic acid extraction on the QIAsymphony SP instrument (Qiagen, Hilden, Germany) using the QIAsymphony® Virus/Bacteria mini kit (cat. no. 931036) according to the manufacturer’s instructions. Total nucleic acid was eluted in 60 μL elution buffer and stored at −20°C until processing.
unpaired student t-test was used to compare the means of two groups. Wilcoxon rank-sum test was used to assess the median between two groups when the data was not normally distributed. Analysis of variance was performed to determine whether the mean CFU of *S. pneumoniae* recovered was different within triplicates of each swab type or across *S. pneumoniae* strains when the experiments were repeated on three different days. A *p* value less than 0.05 was considered as significant.

**Results**

Pneumococcal recovery from mock specimens by culture

Overall, there was a significant difference in the mean CFU of *S. pneumoniae* recovered across all the swab types (*p* < 0.001) and across the three *S. pneumoniae* strains (*p* = 0.012) tested. There was no statistical differences in the mean CFU of *S. pneumoniae* recovered within triplicates of each swab type performed on the same day (*p* = 0.85) or when the experiments were repeated on three different days (*p* = 0.89), suggesting that the experiments were reproducible. As such, the *S. pneumoniae* CFU recovered from triplicates of each swab type and on three different days were pooled.

The percentage recovery of *S. pneumoniae* ATCC 49619 (Serotype 19F) strain from flocked swabs was 100%, while it was 41% from Dacron swabs and 7% from rayon swabs (Table 1). The mean number CFU of *S. pneumoniae* recovered from the ATCC 49619 (serotype 19F) strain using flocked swabs was higher when compared with Dacron (18 × 10⁶ CFU/mL vs. 7.3 × 10⁵ CFU/mL, *p* < 0.001) and rayon swabs (18 × 10⁶ CFU/mL vs. 1.3 × 10⁵ CFU/mL, *p* < 0.001) (Fig. 1A). Dacron swabs released significantly more *S. pneumoniae* (ATCC 49619) than rayon swabs (7.3 × 10⁵ CFU/mL vs. 1.3 × 10⁵ CFU/mL, *p* < 0.001). Similar results were also observed for *S. pneumoniae* serotypes 1 (Fig. 1B) and 5 (Fig. 1C).

Pneumococcal recovery from NP samples collected from healthy children

**Cultivation.** Paired (flocked and Dacron) NP samples were collected from 42 healthy children (median age 8 [IQR 5–16] months) according to the WHO *S. pneumoniae* carriage protocol [28]. *S. pneumoniae* was cultured from 18 of the 42 (43%) NP samples. In 12 of 42 (29%) participants *S. pneumoniae* was recovered from both flocked and Dacron swabs, whilst in four (10%) and two (5%) participants *S. pneumoniae* was recovered only from flocked and Dacron swabs respectively. The children from whom *S. pneumoniae* was recovered were younger than those from whom it was not (8 months vs. 12 months, *p* = 0.032). Although the median number of CFU recovered from flocked swabs was approximately two-fold higher (8.8 × 10⁴ CFU/mL [IQR, 2.0 × 10⁴ – 4.0 × 10⁴ CFU/mL]) than that of Dacron swabs (3.7 × 10⁴ CFU/mL [IQR, 4.0 × 10⁴–3.2 × 10⁴ CFU/mL]), this difference was not significant, *p* = 0.17.

**Quantitative real-time PCR.** Using qPCR, *S. pneumoniae* was detected in 27 of the 42 (64%) NP samples overall, with similar detection rates from flocked and Dacron swabs (26 out of 42 [62%] vs. 24 out of 42 [57%]; *p* = 0.657). *S. pneumoniae* was detected in both flocked and Dacron swabs in 55% (23 out of 42)
Table 1. Recovery of Streptococcus pneumoniae from mock specimens using flocked, Dacron and rayon swabs.

<table>
<thead>
<tr>
<th>S. pneumoniae strain (100% CFU recovery)¹</th>
<th>Swab type</th>
<th>Dilution</th>
<th>n²</th>
<th>Mean CFU recovery/mL (SD)³</th>
<th>Percentage of recovery ⁴</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serotype 19F³ (1.8×10⁶)</td>
<td>flocked</td>
<td>1/1000</td>
<td>9</td>
<td>18×10⁶ (5.7×10⁵)</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Dacron</td>
<td>1/1000</td>
<td>9</td>
<td>7.3×10⁶ (3.7×10⁵)</td>
<td>41</td>
</tr>
<tr>
<td></td>
<td>rayon</td>
<td>1/1000</td>
<td>9</td>
<td>1.3×10⁶ (2.7×10⁵)</td>
<td>7</td>
</tr>
<tr>
<td>Serotype 1 (1.9×10⁶)</td>
<td>flocked</td>
<td>1/1000</td>
<td>9</td>
<td>19×10⁶ (5.4×10⁵)</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Dacron</td>
<td>1/1000</td>
<td>9</td>
<td>8.5×10⁶ (3.8×10⁵)</td>
<td>45</td>
</tr>
<tr>
<td></td>
<td>rayon</td>
<td>1/1000</td>
<td>9</td>
<td>1.0×10⁶ (2.1×10⁵)</td>
<td>5</td>
</tr>
<tr>
<td>Serotype 5 (5.8×10⁷)</td>
<td>flocked</td>
<td>1/1000</td>
<td>9</td>
<td>5.8×10⁷ (3.2×10⁷)</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Dacron</td>
<td>1/1000</td>
<td>9</td>
<td>2.6×10⁷ (8.2×10⁶)</td>
<td>45</td>
</tr>
<tr>
<td></td>
<td>rayon</td>
<td>1/1000</td>
<td>9</td>
<td>0.7×10⁷ (1.1×10⁶)</td>
<td>12</td>
</tr>
</tbody>
</table>

An ototype 19Frb type tested swab type from three the data was determined by (a) as determined by the coefficient of variatSD, Standard deviation. ³ Represents skim milk-tryptone-glucose-glycerol (STGG) media simulating a 100% release of S. pneumoniae genome copies/mL [IQR, 1.3×10⁶–1.8×10⁷] was significantly higher than from Dacron swabs (9.3×10⁷ genome copies/mL [IQR, 7.0×10⁶–1.1×10⁷]; p = 0.005).

The comparison of S. pneumoniae detection by culture and qPCR from NP samples collected from healthy children is summarised in Table 2. The proportion of S. pneumoniae detected by either flocked or Dacron swabs by qPCR was higher when compared to bacterial culture (64% [27/42] vs. 43% [18/42]; p = 0.049).

Discussion

A report from World Health Organisation (WHO) working group [20] and unpublished data from the US Centers for Disease Control and Prevention (CDC) [31] suggest the use of either Dacron or calcium alginate swabs collected in STGG transport medium for S. pneumoniae carriage studies. The present study demonstrated that flocked swabs were better compared to both Dacron and rayon swabs for recovery of S. pneumoniae from mock specimens. In addition, Dacron swabs recovered significantly more S. pneumoniae than rayon swabs. The findings from this study are in line with what has previously been shown for other bacteria [11,21,32]. For instance, Verhoeven et al. showed that flocked swabs were superior to rayon swabs for the recovery of Staphylococcus aureus using culture [20]. Similarly, flocked swabs have been shown to improve the recovery of epithelial cells and viruses compared to rayon swabs [12,13,16]. Our finding, that Dacron swabs were better than rayon swabs for the recovery of pneumococcus is in contrast with the finding of Rubin et al. (2008), following a similar protocol [23]. In our study, we used twice the volume of medium to inoculate swabs (20 µL vs. 10 µL) used by Rubin et al. in their in-vitro assay [23].

Whilst we found no significant statistical difference between flocked and Dacron swabs for recovery of S. pneumoniae by bacterial culture from NP samples collected in children (p = 0.17), significantly greater bacterial load of S. pneumoniae was detected by qPCR from flocked swabs compared to Dacron swabs (p = 0.005). The importance of bacterial load has been recently demonstrated by Albrich et al. showing that the density of NP colonization by S. pneumoniae was higher in patients with pneumococcal community-acquired pneumonia compared to control patients using Dacron swabs for both culture and lytA-targeted qPCR [33]. Further, using the PCR-based methods, the severity of pneumonia caused by S. pneumoniae has been shown to be associated with an increased bacterial load in two independent studies using serum/blood samples [8,34].

The present study did not assess the ability of each swab type tested for collecting other types of specimen, such as sampling of solid surfaces, vaginal and anal specimen collection. However,

Table 2. Comparison of Streptococcus pneumoniae detection by culture and real-time PCR from healthy children.

<table>
<thead>
<tr>
<th>Real-time PCR</th>
<th>Dacron swab</th>
</tr>
</thead>
<tbody>
<tr>
<td>flocked swab</td>
<td>Total</td>
</tr>
<tr>
<td>Positive</td>
<td>16 (38%)</td>
</tr>
<tr>
<td>Negative</td>
<td>10 (24%)</td>
</tr>
<tr>
<td>Total</td>
<td>26 (62%)</td>
</tr>
<tr>
<td>Culture</td>
<td>16 (38%)</td>
</tr>
<tr>
<td>Negative</td>
<td>16 (38%)</td>
</tr>
<tr>
<td>Total</td>
<td>26 (62%)</td>
</tr>
</tbody>
</table>

PCR, Polymerase chain reaction targeting the autolysin gene (lytA) gene of Streptococcus pneumoniae.

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other studies have shown that flocked swabs are more efficient in recovery of bacteria (e.g. Bacillus atrophaeus, Enterococcus hirae) from solid surfaces [11,32,35,36]. In addition, flocked swabs have been shown to collect similar or slightly more cells from anaerobic specimens than Dacron swabs [37]. It has also been shown that the DNA extraction methods may influence the DNA recovery from both cotton and flocked swabs used for the collection of DNA from saliva stains [38]. Therefore, in order to assess whether the observed high bacterial load in this study from flocked swabs is not inherent to the DNA extraction method, studies using different nucleic acid extraction methods are warranted.

In summary, our findings suggest that flocked swabs may offer an improved opportunity for recovery and detection of S. pneumoniae from NP swabs. Importantly, flocked swabs are increasingly used for NP sampling for detection of respiratory viruses by nucleic acid amplification [13,15,39,40]. The use of a single swab type for both virological and bacterial studies would simplify specimen collection protocols [27].

Acknowledgments

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Author Contributions

Conceived and designed the experiments: FD MN MK. Performed the experiments: FD EW. Analyzed the data: FD MK MN HZ EW. Contributed reagents/materials/analysis tools: MN HZ. Wrote the paper: FD MK EW HZ MN.

References

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bacillus spore contaminants from rough surfaces: a challenge to space mission
comparison of dacron versus Flocked nylon swabs for anal cytology specimen
and retrieval from two swab types (cotton and nylon flocked swab) when
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medium and rayon-bud swabs with a sponge reservoir of viral transport medium
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